

Rosaceae/Rose family

## Holodiscus (K. Koch) Maxim.

ocean-spray

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**Growth habit, occurrence, and use.** *Holodiscus* is a taxonomically complex genus and includes about 6 species of western North America and northern South America (Hitchcock and others 1961; Ley 1943). The 2 generally recognized North American species (table 1) are creambush ocean-spray and gland ocean-spray. They are deciduous, multi-stemmed shrubs with simple, alternate, deciduous, toothed to shallowly lobed, exstipulate leaves.

Creambush ocean-spray grows from 1 to 6 m in height, with slender, arching branches and grayish red exfoliating bark. It is a prolific root sprouter, capable of recovering from fire, grazing, or mechanical damage by resprouting from perennating buds in the root crown. Growing at elevations from sea level to 2,150 m, it is most abundant in coastal areas from British Columbia to southwestern California. It also occurs eastward to Montana in drier conifer types of the interior Pacific Northwest. A dominant shrub in a number of forested communities, creambush ocean-spray is also common in riparian areas and on rocky talus slopes (Halversen and others 1986; Topik and others 1986). Remnant stands are found on higher peaks of Great Basin mountain ranges (Hitchcock and others 1961; USDA FS 1937).

Gland ocean-spray is a low, intricately branched shrub that is 0.1 to 3 m tall (Harrington 1954). It differs from creambush ocean-spray in its more compact growth habit, leaves with decurrent petioles, and leaf lobes or teeth without secondary teeth. Gland ocean-spray grows east of the Cascade Mountains and the Sierra Nevada, from north central Oregon to Chihuahua, Mexico, at elevations ranging from 1,400 to 3,350 m (Harrington 1954; Mozingo 1987; USDA FS 1937). Although gland ocean-spray is found in a variety of plant communities, its most characteristic habitats are talus slopes, rock outcrops, slickrock plateaus, and dry, rocky desert areas.

Palatability and forage value of both ocean-spray species vary geographically but are generally low for livestock and big game. However, in the absence of more palatable shrubs, substantial quantities are browsed by deer and elk on low-elevation winter ranges. In some areas, ocean-sprays are important year-round (USDA FS 1937). Both shrubs may increase on summer ranges where other forage species are browsed preferentially (Ferguson 1983). Gland ocean-spray is browsed in summer by bighorn sheep and both species are browsed by rabbits (Sutton and Johnson 1974; Todd 1975; van Dersal 1938).

Ocean-spray has considerable potential for re-vegetating a variety of disturbed areas. Populations capable of growing on dry, rocky, well-drained sites may be particularly useful (Stark 1966; Sutton and Johnson 1974). Ocean-spray has been recommended for use in nonintensive highway plantings, riparian areas, windbreaks, erosion control projects, wildlife habitat improvement projects, and conservation plantings (Antieau 1987; Athowe 1993; Flessner and others 1992). Because of their growth habits, showy inflorescences, and fall coloration, both species are attractive ornamentals. Creambush ocean-spray was first cultivated in 1827 and gland ocean-spray in 1853 (Rehder 1940).

Native Americans made digging sticks and arrow shafts from the hard wood and straight branches of ocean-spray (Anderson and Holmgren 1969; Daubenmire 1970; Hopkins and Kovalchik 1983). Fruits of gland ocean-spray were eaten by Native Americans of the Great Basin, and pioneers made nails from its wood.

Both North American ocean-sprays are tetraploid, with  $2X = n = 18$  (Antieau 1986; Goldblatt 1979; McArthur and Sanderson 1985), and both exhibit considerable morphological variation. A genetic basis for variability in such characteristics as growth habit, growth rate, leaf morphology, and flower abundance in creambush ocean-spray is suggested by common garden studies (Flessner and others 1992).

**Flowering and fruiting.** Although the showy terminal panicles and floral buds of both species develop in early spring, flowering is delayed until late spring to mid-summer. Fruits ripen in late summer and are dispersed by wind and gravity through November (Hitchcock and others 1961; Stickney 1974) (table 2). The insect-pollinated flowers are small, creamy-white, perfect, and perigynous (Hitchcock and others 1961; McArthur 1984). The entire disk lining the hypanthium gives the genus its name (Greek: *holo* = whole and *diskos* = disk). Each flower produces 5 villous, light-yellow achenes that are about 2 mm long (figures 1 and 2). Seeds are broadly oblong and contain a thin endosperm and an embryo with ovate cotyledons (Ley 1943) (figure 1).

**Collection, cleaning, and storage.** Ocean-spray achenes are among the smallest of shrub fruits. Estimates of the number of cleaned achenes per weight exceed 11,000,000/kg (5,000,000/lb) for each species (King 1947; Link 1993). Achene collection is tedious, and supplies are rare and costly. In addition, the achenes are difficult to handle because of their pubescence and small size. Achenes are hand-stripped from inflorescences in late summer or autumn (table 2) (Monsen 1996). Large debris in air-dried collections can be removed with a fanning mill. Small lots may be cleaned by hand-rubbing and sieving (Link 1993).

Sound achenes are identified by examining imbibed achenes through a dissecting microscope for the presence of an embryo. Using this method, King (1947) found that only 7% of ocean-spray achenes collected were sound. In creambush ocean-spray seeds collected in northern Idaho, only 7% were sound. In creambush ocean-spray from British Columbia, viability was greater for achense collected in October or November than for those collected in August or September (Marchant and Sherlock 1984).

Storage requirements for ocean-spray have not been examined. The achenes appear to be orthodox in storage behavior and can probably be stored for several years at low water contents and temperatures.

**Germination.** Germination of creambush ocean-spray seeds is enhanced by wet

prechilling at 2 to 5 °C for 15 to 18 weeks (King 1947; Marchant and Sherlock 1984). King (1947) obtained 84% germination in 22 days when seeds were prechilled for 18 weeks before incubation at 20 to 24 °C.

Germination of gland ocean-spray has received little study. There are no official testing prescriptions for this genus, and effective pretreatments have not been reported. Link (1993) reported that 16 weeks of wet prechilling failed to release dormancy in this species. Effective treatments have not been reported.

Viability of ocean-spray seeds may be tested by tetrazolium chloride staining. After 3 hours of imbibition in water at room temperature, seeds are excised from the achene and the seedcoat is pricked or slit near the center of the seed. Seeds are then imbibed in 1% tetrazolium chloride for 4 hours at room temperature. Stained embryos may be read in place, as the seedcoat is very thin (Hurd 1996; King 1947). Staining should be evaluated as described by Grabe (1970) for dicotyledonous seeds other than legumes.

**Nursery practice.** Ocean-sprays may be propagated as bareroot or container stock (Everett 1957). Achenes should be fall-sown or artificially prechilled and spring-sown in bareroot nurseries (Flessner and others 1992). Marchant and Sherlock (1984) obtained successful plantings only when freshly harvested achenes were planted in fall. Cleaned achenes of both species can be drilled at reasonably uniform spacings within rows (Shaw and Monsen in press). They may also be broadcast and covered by dragging a lightweight chain over the seedbed. Seedlings develop slowly and may be lifted as 1+0 or 2+0 stock, depending upon size specifications and growing conditions.

Container seedlings are propagated by planting several wet-prechilled achenes in each container and thinning or by planting germinants. Kruckeberg (1982) reported that ocean-spray can be propagated by fall-sowing achenes in boxes outdoors and covering them lightly with soil. Flessner and others (1992) planted wet prechilled (4 months at 4 °C) creambush ocean-spray achenes in shallow flats in a greenhouse. Seedlings emerged after 16 to 30 days of incubation at a minimum temperature of 21 °C. Developing seedlings were fertilized and treated with a fungicide as necessary. After 2 months they were transferred to larger containers in a lathhouse where they were held overwinter for planting as 1+0 stock.

Kruckeberg (1982) reported that creambush ocean-spray planting stock is easily obtained by potting up wildlings, which he found were often abundant. Morgan and Neuenschwander (1988) observed high densities of creambush ocean-spray wildlings following severe burns, but Wright and others (1979) and Stickney (1996) concluded that the species exhibits poor seedling regeneration following fire in sagebrush and conifer communities of the Intermountain and northern Rocky Mountain regions.

Ocean-spray can be grown from cuttings, but rooting of both species varies widely among clones, cutting types, and propagation techniques (Antieau 1987; Link 1993). Softwood cuttings may be treated with rooting hormones and propagated in a greenhouse with a mist system (Antieau 1987; Marchant and Sherlock 1984). Success with semi-hardwood cuttings has been variable (Everett and others 1978; Kruckeberg 1982). Fall-harvested hardwood cuttings are cut to 15-cm (6-in) lengths and treated with 0.8% indole-3-butyric acid (IBA) powder and a fungicide (Macdonald 1986). Hardwood cuttings stored in straw-bale bins or cold frames will develop

calluses (Macdonald 1986; Marchant and Sherlock 1984). When fall-planted, these cuttings root rapidly. Layers and suckers have been propagated successfully (Kruckeberg 1982).

**Field practice.** Fresh achenes broadcast over a rough seedbed in fall are covered by natural soil sloughing (Shaw and Monsen in press; Van Dersal 1938). Achenes may be mixed with seeds of other shrub species, but they should not be sown with more competitive grasses or forbs. Planting areas should be selected carefully to make the best use of seed supplies, as seeding results are often erratic. Native creambush ocean-spray seedlings develop slowly and are poor competitors (Wright and others 1979).

Creambush ocean-spray can be established by transplanting. Youtie (1992) reported good survival of rooted cuttings on biscuit scablands in Oregon's Columbia River Gorge. Marchant and Sherlock (1984) found that planted seedlings grew slowly the first year. Low survival on western Montana roadcuts was attributed to poor soils and unhealthy planting stock (Hungerford 1984).

### References

- Anderson BA, Holmgren AH. 1969. Mountain plants of northeastern Utah. Circ. 319. Logan: Utah State University. 148 p.
- Antieau CJ. 1986. Patterns of natural variation in ocean-spray (*Holodiscus discolor*) (Rosaceae). HortScience 21: 120.
- Antieau CJ. 1987. Field notes: *Holodiscus discolor*. American Nurseryman 166: 110.
- Atthowe H. 1993. Propagation of riparian and wetland plants. In: Landis TD, tech. coord. Proceedings, Western Forest Nursery Association; 1992 September 14B18; Fallen Leaf Lake, CA: Gen. Tech. Rep. RM-221. Fort Collins, CO: USDA Forest Service, Rocky Mountain Forest and Range Experiment Station: 78B81.
- Daubenmire R. 1970. Steppe vegetation of Washington. Tech. Bull. 62. Pullman: Washington State University.
- Drew LA. 1967. Comparative phenology of seral shrub communities in the cedar/hemlock zone [thesis]. Moscow, ID: University of Idaho. 108 p.
- Everett PC. 1957. A summary of the culture of California plants at the Rancho Santa Ana Botanic Garden. Claremont, CA: Rancho Santa Ana Botanic Garden. 223 p.
- Everett RL, Meeuwig RO, Robertson JH. 1978. Propagation of Nevada shrubs by stem cuttings. Journal of Range Management 31: 426B429.
- Ferguson RB. 1983. Use of rosaceous shrubs for wildland plantings in the Intermountain West. In: Monsen SB, Shaw N, comps. Proceedings; Managing Intermountain Rangelands: Improvement of Range and Wildlife Habitats; 1981 September 15B17; Twin Falls, ID, & 1982 June 22B24; Elko, NV: Gen. Tech. Rep. INT-157. Ogden, UT: USDA Forest Service, Intermountain Forest and Range Experiment Station: 136B139.
- Flessner TR, Darris DC, Lambert SC. 1992. Seed source evaluation of four native riparian shrubs for streambank rehabilitation in the Pacific Northwest. In: Clary WP, McArthur ED, Bedunah D, Wambolt CL, comps. Proceedings, Symposium on Ecology and Management of Riparian Shrub Communities; 1991 May 29B31; Sun Valley, ID: Gen. Tech. Rep. INT-289. Ogden, UT: USDA Forest Service, Intermountain Research Station:

- 155B162.
- Goldblatt P. 1979. Miscellaneous chromosome counts in Angiosperms: 2. Including new family and generic records. *Annals of Missouri Botanic Gardens* 66: 856B861.
- Grabe DF, ed. 1970. Tetrazolium testing handbook for agricultural seeds. In: Handbook on seed testing. *Contrib. 29. Association of Official Seed Analysts*. 62 p.
- Halverson NM, Topik C, Van Vickle R. 1986. Plant association and management guide for the western hemlock zone: Mt. Hood National Forest. R6-ECOL-232A. Portland, OR: USDA Forest Service, Pacific Northwest Region. 111 p.
- Harrington HD. 1954. Manual of the plants of Colorado. Denver: Sage Books. 666 p.
- Hitchcock CL, Cronquist A, Ownbey M, Thompson JW. 1961. Vascular plants of the Pacific Northwest: 3. Saxifragaceae to Ericaceae. Seattle: University of Washington Press. 614 p.
- Hopkins WE, Kovalchik BL. 1983. Plant associations of the Crooked River National Grassland. R6 Ecol. 133-1983. Portland, OR: USDA Forest Service, Pacific Northwest Region. 98 p.
- Hungerford RD. 1984. Native shrubs: suitability for revegetating roadcuts in northwestern Montana. Res. Pap. INT-331. Ogden, UT: USDA Forest Service, Intermountain Forest and Range Experiment Station. 13 p.
- Hurd EG. 1996. Unpublished data. Boise, ID: USDA Forest Service, Intermountain Research Station.
- Jorgensen K. [in press]. Appendix I. In: Monsen SB, Stevens R, comps. Restoring western ranges and wildlands. RM-GTR-XX. Fort Collins, CO: USDA Forest Service, Rocky Mountain Research Station.
- King JE. 1947. The effect of various treatments to induce germination of seeds of some plants valuable for soil conservation and wildlife [thesis]. Moscow, ID: University of Idaho. 97 p.
- Kruckeberg, AR. 1982. Gardening with native plants of the Pacific Northwest. Seattle: University of Washington Press. 252 p.
- Ley A. 1943. A taxonomic revision of the genus *Holodiscus* (Rosaceae). *Bulletin of the Torrey Botanical Club* 70: 275B288.
- Link E. 1993. Native plant propagation techniques for National Parks: interim guide. East Lansing, MI: Rose Lake Plant Materials Center. 240 p.
- Macdonald B. 1986. Practical woody plant propagation for nursery growers. Volume 1. Portland, OR: Timber Press. 669 p.
- Marchant C, Sherlock J. 1984. A guide to selection and propagation of some native woody species for land rehabilitation in British Columbia. For. Res. Rep. RR84007-HQ. Victoria, BC: British Columbia Ministry of Forests. 117 p.
- McArthur ED. 1984. Natural diversity of western range shrubs. In: Cooley JL, Cooley JH, eds. Natural diversity in forest ecosystems. Proceedings, 1982 November 29-December 1; Athens, GA. Athens, GA: University of Georgia, Institute of Ecology: 193B209.
- McArthur ED, Sanderson SC. 1985. A cytotoxic contribution to the western North American rosaceous flora. *Madroño* 32: 24B28.
- McMurray NE. 1987a. *Holodiscus discolor*. In Fischer WC, comp. The Fire Effects Information System [database on magnetic tape]. Missoula, MT: USDA Forest Service,

- Intermountain Research Station, Intermountain Fire Sciences Laboratory.
- McMurray NE. 1987b. *Holodiscus dumosus*. In Fischer WC, comp. The Fire Effects Information System [database on magnetic tape]. Missoula, MT: USDA Forest Service, Intermountain Research Station, Intermountain Fire Sciences Laboratory.
- Monsen SB. 1996. Unpublished data. Provo, UT: USDA Forest Service, Rocky Mountain Research Station.
- Morgan P, Neuenschwander LF. 1988. Seed-bank contributions to regeneration of shrub species after clear-cutting and burning. Canadian Journal of Botany 66: 169B172.
- Mozingo HN. 1987. Shrubs of the Great Basin. Reno: University of Nevada Press. 342 p.
- Munz PA, Keck DD. 1973. A California flora. Berkeley: University of California Press. 1681 p.
- Orme ML, Leege TA. 1980. Phenology of shrubs on a north Idaho elk range. Northwest Science 54: 187B198.
- Rehder A. 1940. Manual of cultivated trees and shrubs. 2nd ed. New York: MacMillan. 996 p.
- Shaw NL, Monsen SB. [in press]. Rosaceous shrubs. In: Monsen SB, Stevens R, comps. Restoring western ranges and wildlands. RM-GTR-XX. Fort Collins, CO: USDA Forest Service, Rocky Mountain Research Station.
- Stark N. 1966. Review of highway planting information appropriate to Nevada. Coll. Agric. Bull. B-7. Reno: University of Nevada, Desert Research Institute. 209 p.
- Stickney PF. 1974. *Holodiscus discolor* (Pursh) Maxim., creambush rockspiraea. In: Shoptmeyer, CS, tech. coord. Seeds of woody plants in the United States. USDA Agric. Handb. 450. Washington, DC: USDA Forest Service: 448B449.
- Stickney PF. 1996. Personal communication. Missoula, MT: USDA Forest Service, Rocky Mountain Research Station.
- Sutton R, Johnson CW. 1974. Landscape plants from Utah's mountains. EC-368. Logan: Utah State University. 137 p.
- Todd JW. 1975. Foods of Rocky Mountain bighorn sheep in southern Colorado. Journal of Wildlife Management 39: 108B111.
- Topik C, Halverson NM, Brockway DG. 1986. Plant association and management guide for the western hemlock zone: Gifford Pichot National Forest. R6-ECOL-230A. Portland, OR: USDA Forest Service, Pacific Northwest Region. 132 p.
- USDA FS [USDA Forest Service]. 1937. Range plant handbook. Washington, DC. 512 p.
- Van Dersal WR. 1938. Native woody plants of the United States: their erosion control and wildlife values. Misc. Pub. 303. Washington, DC: U.S. Department of Agriculture. 362 p.
- Welsh SL, Atwood ND, Higgins LC, Goodrich S. 1987. A Utah flora. Great Basin Naturalist 9: 1B894.
- Wright HA, Neuenschwander LF, Britton CM. 1979. The role and use of fire in sagebrush-grass and pinyon-juniper plant communities: a state of the art review. Gen. Tech. Rep. INT-58. Ogden, UT: USDA Forest Service, Intermountain Forest and Range Experiment Station. 48 p.
- Youtie BA. 1992. Biscuit scabland restoration includes propagation studies (Oregon). Restoration & Management Notes 10: 79B80.

**Figure 1** *Holodiscus*, ocean spray: achenes.

**Figure 2** *Holodiscus dumosus*, gland ocean-spray: longitudinal section through an achene.

**Figure 3** *Holodiscus dumosus*, gland ocean-spray: seedling development.